

Addendum

The BnALMT1 Protein that is an Aluminum-Activated Malate Transporter is Localized in the Plasma Membrane

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KEY WORDS

Brassica napus, Aluminum toxicity, malate efflux, BnALMT, GFP, PM, *Nicotiana tabacum*, *Xenopus laevis* oocytes and Al-tolerance

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Addendum to:

The BnALMT1 and BnALMT2 Genes from Rape Encode Aluminum-Activated Malate Transporters That Enhance the Aluminum Resistance of Plant Cells

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ABSTRACT

We have previously reported that Al-induces citrate and malate efflux from P-sufficient and P-deficient plants of rape (*Brassica napus* L.) and that P-deficiency alone could not induce this response. Further investigation showed that the transcript of two genes designated BnALMT1 and BnALMT2 is accumulated in roots by Al-treatment. Transgenic tobacco cells (*Nicotiana tabacum*) and *Xenopus laevis* oocytes expressing the BnALMT1 and BnALMT2 proteins released more malate than control cells in the presence of Al, indicating that the BnALMT genes encode an Al-activated malate transporter. The transgenic tobacco cells exposed to toxic level of Al grew better than control cells indicating that the genes can enhance Al-resistance of plant cells. In this study we showed the subcellular localization of BnALMT1 fused to the green fluorescence protein (GFP). The BnALMT1::GFP construct was transiently expressed in protoplasts prepared from Arabidopsis leaves using the polyethylene glycol (PEG) method. The result showed that the BnALMT1 protein is localized in the plasma membrane. This provides further evidence that the BnALMT proteins facilitate the transport of malate across the plasma membrane (PM).

Aluminium toxicity limits crop production on the acidic soils that comprise over 40% of the world's potentially arable lands.¹ On such soils, P-deficiency is commonly reported together with Al-toxicity.² When the soil pH falls below 5, Al is solubilized into toxic forms and then inhibits root growth and functions.

The release of organic anions such as malate, citrate and oxalate from plant roots is believed to be a mechanism of Al resistance.³⁻⁸ Induction of organic anions efflux was also reported in P-deficient rape,⁹ white lupin¹⁰ and alfalfa.¹¹ However, in our previous investigation we did not observe any induction of organic anion efflux under P-deficient condition,¹² but we have shown that Al-induced efflux is more pronounced in P-sufficient plants of rape.

The Al resistance in wheat relies on the Al-dependent efflux of malate anions from the root apices and a strong correlation exists between relative Al resistance of different genotypes and the capacity for malate efflux.^{4,13} Pharmacological and electrophysiological studies suggest that malate efflux is facilitated by anion channels and this is consistent with the electrochemical gradient for anions across the plasma membrane.^{14,15}

The *TaALMT1* gene which controls the Al-dependent efflux of malate was first isolated from wheat.¹⁶ *TaALMT1* expression in Al-resistant genotypes of wheat is 5 to 10-fold higher than in Al-sensitive genotypes.^{16,17} Heterologous expression of *TaALMT1* in cultured-tobacco cells, *Xenopus* oocytes, and intact rice and barley plants conferred an Al-activated malate efflux and increased the Al-tolerance of tobacco suspension cells and intact barley plants.^{16,18} Recently *AtALMT1*, one of the homologues of *TaALMT1* in Arabidopsis, has been implicated in an Al-resistance mechanism which also relies on malate release.¹⁹ In an effort to elucidate the molecular mechanism underlying the Al-induced malate efflux in rape we have cloned two homologues of the *TaALMT1* gene, designated *BnALMT1* and *BnALMT2*. The transcript of *BnALMT1* and *BnALMT2* was accumulated in the root, not shoot by Al in a concentration dependant manner.²⁰ This is similar to *AtALMT1*¹⁹ but contrasts with *TaALMT1* which is constitutively expressed in wheat and unaffected by Al treatment.¹⁶ Our result also showed that neither *BnALMT1* nor *BnALMT2* is induced by P-deficiency.²⁰ Therefore, we conclude that P-deficiency does not induce both *BnALMT* transcript accumulation and malate efflux in rape.^{12,20}

Overexpression of the BnALMT proteins in tobacco cells and *Xenopus* oocytes conferred an Al-induced malate efflux. Citrate was not released from the transgenic cells, indicating that the BnALMT proteins are specific for malate transport. Hence, we suggest that other proteins might be involved in facilitating the Al-induced citrate efflux from wild

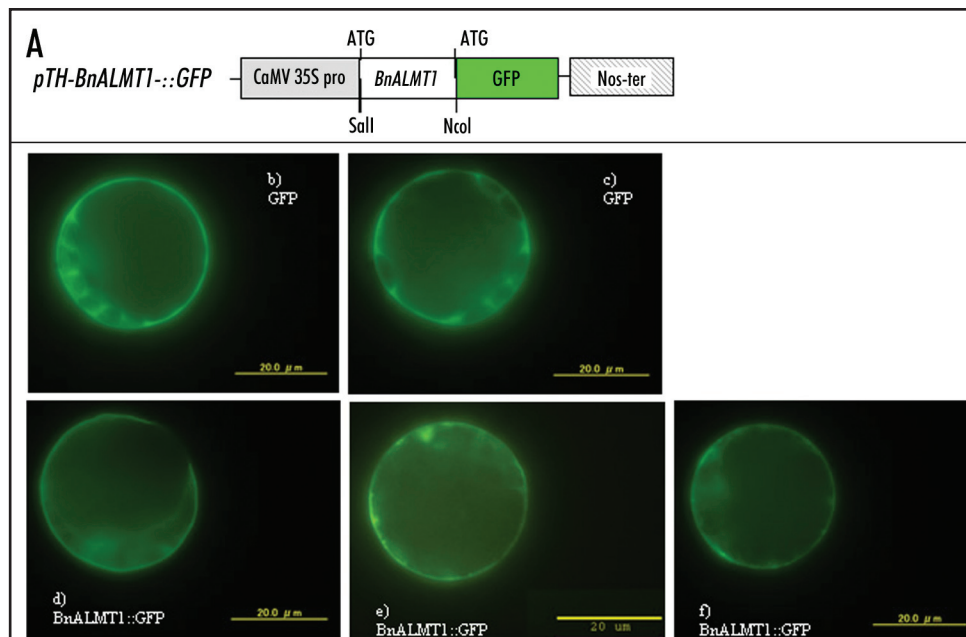


Figure 1. Expression of BnALMT1::GFP fusion protein in Arabidopsis (ecotype Colombia) leaf protoplast. BnALMT1 was cloned into pTH-2 at the Sall and NcoI site in frame with the GFP on the C-terminal site to yield a construct pTH-BnALMT1::GFP (A). The construct was transiently expressed in Arabidopsis leaf protoplast using the PEG method of transformation (D–F). For comparison, pTH-GFP was used as control (B) and (C). Representative images are presented.

type plants of rape.¹² In line with this, other researchers have identified an Al tolerance locus in sorghum that segregates with citrate release from roots.²¹ If such genes are identified, future studies can examine whether similar genes are involved in citrate release from rape. Furthermore, heterologous expression of the BnALMT proteins enhanced the Al-resistance of the transgenic tobacco cells. It is worthwhile therefore to overexpress the BnALMT proteins in other economically important crops in order to maximize crop productivity on the acidic soils where Al-toxicity is a major problem.

The present study focused on the subcellular localization of BnALMT1 protein. For this purpose, BnALMT1 was cloned into pTH-2 vector at the Sall and NcoI site in frame with the green fluorescence protein (GFP) fused to the C-terminal site (Fig. 1a). The construct under the control of 35S promoter and nopaline synthase (nos) terminator was transiently expressed in Arabidopsis leaf protoplast using the polyethylene glycol (PEG) method as previously described.²² Transformed protoplasts were examined using a fluorescent microscope (BX61 equipped with a sectioning device DSU, Olympus) with a B-excitation filter set (for GFP).

The result showed that the fluorescent signal of the BnALMT1::GFP construct was more confined to the plasma membrane (Figs. 1d, 1e and 1f) unlike the control (expressing only the GFP) which seems not to be restricted to the PM but also observed in the cytoplasm (Figs. 1b and 1c). Similarly, the TaALMT1 protein is localized in the plasma membrane.²³ Therefore, we conclude that the BnALMT proteins are localized in the plasma membrane and facilitate the transport of malate out of plant cells exposed to toxic Al.

The responses of rape genotypes to Al-stress have not been well understood. Hence, future study will examine these responses and whether genotypic variation in Al-tolerance exists among rape genotypes. Furthermore, the polymorphism in the two *BnALMT1* genes can be studied to generate a molecular marker which can be used to map the location of the two genes on the rape genome. If significant

variations are observed among the parental lines, the marker can be used to study if Al-tolerance segregates with either of the *BnALMT* genes.

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